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## 1 FIGURE 9 – BACCALA ET AL. (2016) DTF: UNIFIED ASYMPTOTIC THEORY

DESCRIPTION:

Routine **figure9\_example3\_idtf\_ns2000.m** publish

Linear five-dimensional VAR(2) model

LA Baccala, DY Takahashi, K Sameshima (2016) Directed Transfer Function: Unified Asymptotic Theory and Some of its Implications. *IEEE Transactions on Biomedical Engineering* **PP**.

<http://dx.doi.org/10.1109/TBME.2016.2550199>

Example 3: Asymptotic DTF: Example 3 5-VAR model - open / closed loop.

```
x1 ==> x2 ==> x3 ==> x4 <==> x5
^-----/
```

```
*This is the close loop model.*
```

## Contents

- Generating data set for analysis
- Equation (11)
- Connectivity diagram
- DTF estimation
- Figure depicted in the article Baccala et al (2016)
- Some remarks:

## Generating data set for analysis

```
clear; clc; format compact
flgPlotStyle = 'Print'; % or 'Screen' mode
flgRandomize = 0; % Generate the specific data set used in Fig. 9.
ns = 2000; % number of sample points
nDiscard = 20000; % number of points discarded at beginning of simulation
p = 2; % model order

if (exist('figure9_example3_idtf_ns2000.mat') == 2) & is_octave & ~flgRandomize
    load figure9_example3_idtf_ns2000
else
    [u] = fbaccala2016_example3( ns, nDiscard, flgRandomize );
    if ~is_octave & ~flgRandomize,
```

```

    save figure9_example3_idtf_ns2000 u
end;
end;

chLabels = []; % Using default labeling schema for channel identification

```

```

=====
Asymptotic DTF: Example 3 5-VAR model - open / closed loop.
x1 ==> x2 ==> x3 ==> x4 <==> x5
^-----/
This is the close loop model.
=====

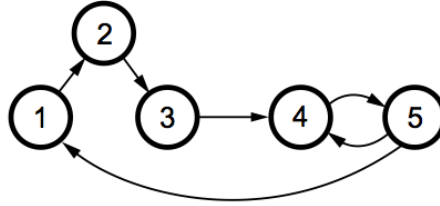
```

Equation (11)

$$\begin{aligned}
 x_1(n) &= 0.25\sqrt{2}x_1(n-1) + 0.25\sqrt{2}x_2(n-1) + w_1(n) \\
 x_2(n) &= -0.25\sqrt{2}x_1(n-1) + 0.25\sqrt{2}x_2(n-1) + w_2(n) \\
 x_3(n) &= 0.5x_2(n-1) + w_3(n)
 \end{aligned} \tag{12}$$

Equation (11) from Baccala et al. *IEEE Trans Biomed Engin.*, 2016.

Connectivity diagram



Example 3 loop connectivity structure following (11). Signals from any structure reach all other structures. from Baccala et al. *IEEE Trans Biomed Engin.*, 2016.

Data pre-processing: detrending and normalization options

```

flgDetrend = 1; % Detrending the data set
flgStandardize = 0; % No standardization
[nChannels, nSegLength] = size(u);
if nChannels > nSegLength, u = u.';
    [nChannels, nSegLength] = size(u);
end;
if flgDetrend,
    for i=1:nChannels, u(i,:) = detrend(u(i,:)); end;
    disp('Time series were detrended.');
```

Time series were detrended.

## MVAR model estimation

```
maxIP = 30;           % maximum model order to consider.
alg = 1;              % 1: Nutall-Strand MVAR estimation algorithm
criterion = 1;        % 1: AIC, Akaike Information Criteria
disp('Running MVAR estimation and GCT analysis routines.')
[IP,pf,A,pb,B,ef,eb,vaic,Vaicv] = mvar(u,maxIP,alg,criterion);
disp(['Number of channels = ' int2str(nChannels) ' with ' ...
      int2str(nSegLength) ' data points; MAR model order = ' int2str(IP) ' .']);
```

```
Running MVAR estimation and GCT analysis routines.
maxOrder limited to 30
IP=1  vaic=79027.676838
IP=2  vaic=76136.019225
IP=3  vaic=76162.273276
```

Number of channels = 5 with 2000 data points; MAR model order = 2.

## Testing for adequacy of MAR model fitting through Portmanteau test

```
h = 20; % testing lag
MVARadequacy_signif = 0.05; % VAR model estimation adequacy significance
                        % level
aValueMVAR = 1 - MVARadequacy_signif; % Confidence value for the testing
flgPrintResults = 1;
```

Granger causality test (GCT) and instantaneous GCT

```
gct_signif = 0.01; % Granger causality test significance level
igct_signif = 0.01; % Instantaneous GCT significance level
flgPrintResults = 1;
[Tr_gct, pValue_gct, Tr_igct, pValue_igct] = gct_alg(u,A,pf,gct_signif, ...
                                                       igct_signif,flgPrintResults);
```

```
-----
GRANGER CAUSALITY TEST
=====
Connectivity matrix:
      NaN      0      0      0      1.00
      1.00     NaN      0      0      0
      0      1.00     NaN      0      0
      0      0      1.00     NaN     1.00
      0      0      0      1.00     NaN
Granger causality test p-values:
      NaN      0.54      0.11      0.47      0
      0      NaN      0.27      0.59      0.97
      0.45      0      NaN      0.23      0.88
      0.13      0.89      0      NaN      0
      0.36      0.87      0.49      0      NaN
-----
INSTANTANEOUS GRANGER CAUSALITY TEST
=====
Instantaneous connectivity matrix:
      NaN      0      0      0      0
```

```

0      NaN      0      0      0
0      0      NaN      0      0
0      0      0      NaN      0
0      0      0      0      NaN
Instantaneous Granger causality test p-values:
NaN      0.56      0.64      0.08      0.41
0.56      NaN      0.94      0.33      0.28
0.64      0.94      NaN      0.74      0.03
0.08      0.33      0.74      NaN      0.71
0.41      0.28      0.03      0.71      NaN
>>>> Instantaneous Granger causality NOT detected.

```

## DTF estimation

DTF analysis results are saved in **c** structure. See `asypm_dtf.m` or issue `>> help asypm_dtf` command for more detail.

```

metric = 'info'; % euc = original PDC or DTF;
           % diag = generalized PDC (gPDC) or directed coherence (DC);
           % info = information PDC (iPDC) or iDTF.

nFreqs = 128;
alpha = 0.01;

c = asypm_dtf(u,A,pf,nFreqs,metric,alpha);

```

```
* Information DTF and asymptotic statistics
```

$|_i DTF(\lambda)|^2$  Matrix Layout Plotting

```

switch lower(flagPlotStyle)
case 'print'
    flgColor = [0]; % white background
    flgMax = 'TCI';
    flgSignifColor = 1; % black + gray
    flgScale = 3; % [0 max(flagMax)]
otherwise % 'screen'
    flgColor = [1]; % Colored background
    flgMax = 'TCI';
    flgSignifColor = 3; % red + green
    flgScale = 2; % [0 1]/[0 .1]/[0 .01]
end;

% -----Plotting options flag setting-----
% [1 2 3 4 5 6 7]
flagPrinting=[1 1 1 2 2 0 1];
% | | | | | 7 Spectra(0: w/o SS; 1: Linear; 2: log-scale)
% | | | | | 6 Coherence
% | | | | 5 Plot lower confidence limit (legacy)
% | | | 4 Plot upper confidence limit
% | | 3 Significant DTF(w) in red line (legacy)
% | 2 Patnaik threshold level in black dashed-line
% 1 plot DTF
% -----

fs = 1; % sampling frequency

```

```

w = fs*(0:(nFreqs-1))/2/nFreqs;
w_max = fs/2;

h=figure;
set(h,'NumberTitle','off','MenuBar','none', ...
    'Name','[Asymptotic DTF] Fig 9. Example 3 - iDTF, ns = 2000')

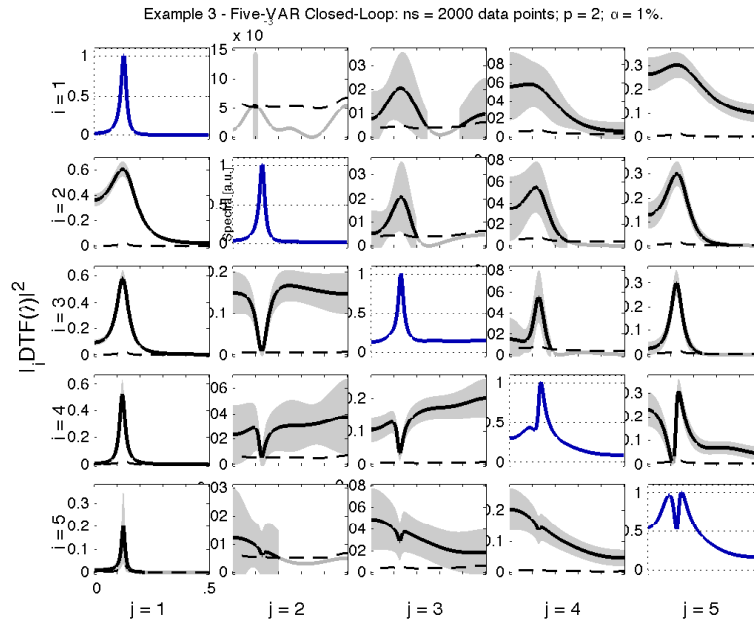
[hxlabel hylabel] = xplot(c,...
    flgPrinting,fs,w_max,chLabels,flgColor,flgScale,flgMax,flgSignifColor);

%xplot_title(alpha, metric, measure(c));

[ax,hT]=suplabel(['Example 3 - Five-VAR Closed-Loop: ns = ' ...
    int2str(ns) ' data points; p = ' int2str(c.p) '; \alpha = ' ...
    int2str(100*alpha) '%.'], 't');
set(hT,'FontSize',10); % Subtitle font size

drawnow

```



Uncomment the command line bellow to generate an eps output file

```
% print -depsc2 -painters Fig9_example3_idtf_ns2000.eps
```

**Figure depicted in the article Baccala et al (2016)**

Figure 9, reproduced from article.

**Some remarks:**

1. As usual, figure 9 underwent some cosmetic edit and addition of a inlet graph

This completes the **Figure 9** generation (Baccala et al, 2016)'

